

UCSF

UC San Francisco Previously Published Works

Title

Thrombin-induced events in non-platelet cells are mediated by the unique proteolytic mechanism established for the cloned platelet thrombin receptor.

Permalink

<https://escholarship.org/uc/item/3zq2b49v>

Journal

The Journal of cell biology, 116(3)

ISSN

0021-9525

Authors

Hung, DT
Vu, TH
Nelken, NA
et al.

Publication Date

1992-02-01

DOI

10.1083/jcb.116.3.827

Peer reviewed

Thrombin-induced Events in Non-Platelet Cells Are Mediated by the Unique Proteolytic Mechanism Established for the Cloned Platelet Thrombin Receptor

David T. Hung,*† Thien-Khai H. Vu,† Nicolas A. Nelken,‡§ and Shaun R. Coughlin*||

Departments of *Laboratory Medicine, †Cardiovascular Research Institute, ‡Surgery, and ||Medicine, University of California at San Francisco, San Francisco, California 94143-0524

Abstract. We recently isolated a cDNA clone encoding a functional platelet thrombin receptor that defined a unique mechanism of receptor activation. Thrombin cleaves its receptor's extracellular amino terminal extension, unmasking a new amino terminus that functions as a tethered peptide ligand and activates the receptor. A novel peptide mimicking this new amino terminus was a full agonist for platelet secretion and aggregation, suggesting that this unusual mechanism accounts for platelet activation by thrombin. Does this mechanism also mediate thrombin's assorted actions on non-platelet cells? We now report that the novel

thrombin receptor agonist peptide reproduces thrombin-induced events (specifically, phosphoinositide hydrolysis and mitogenesis) in CCL-39 hamster lung fibroblasts, a naturally thrombin-responsive cell line. Moreover, these thrombin-induced events could be recapitulated in CV-1 cells, normally poorly responsive to thrombin, after transfection with human platelet thrombin receptor cDNA. Our data show that important thrombin-induced cellular events are mediated by the same unusual mechanism of receptor activation in both platelets and fibroblasts, very likely via the same or very similar receptors.

WE recently isolated a cDNA clone encoding a functional thrombin receptor by expressing cloning in *Xenopus* oocytes (23). The library used was made from Dami cells, a megakaryocyte-like cell line, and the clone was shown to be expressed by platelets. The clone encodes a seven transmembrane domain receptor with a one hundred-residue extracellular amino terminal extension. This extracellular extension contains a putative thrombin cleavage site (LDPR/S; / represents the point of cleavage) resembling the known thrombin cleavage site found in the thrombin-activated zymogen protein C (LDPR/I). Structure-activity studies with the cloned receptor (10, 22, 23) strongly suggest a unique mechanism of receptor activation. Thrombin cleaves its receptor at the LDPR/S cleavage site unmasking a new amino terminus beginning with the sequence SFL...; this new amino terminus then functions as a tethered peptide ligand, binding to an as yet undefined site in the body of the thrombin receptor, effecting receptor activation. A peptide mimicking this new amino terminus was a full agonist for platelet secretion and aggregation, supporting the model of receptor activation proposed above and suggesting that this unusual mechanism accounts for platelet activation by thrombin (22).

In addition to its critical role in activating platelets to effect hemostasis and thrombosis (8), thrombin has potent actions on a variety of non-platelet cell types (reviewed in 16). For example, thrombin has potent chemotactic activities for monocytes (1) and is mitogenic for several cell types (3, 4),

actions that may play important roles in inflammatory and proliferative processes in vivo. Are these assorted activities of thrombin also mediated by the unusual receptor activation mechanism described above? Do multiple receptors mediate thrombin's diverse actions, or does the cloned "platelet thrombin receptor" also account for thrombin-induced events in non-platelet cell types? To begin to address these questions, we took two approaches. First, the hamster lung fibroblast cell line CCL-39 is known to respond to thrombin with robust activation of the phosphoinositide turnover signaling pathway and with mitogenesis (11, 14, 15). We asked whether the novel thrombin receptor agonist peptide described above would elicit these two important thrombin-induced events in this naturally thrombin-responsive cell line. As implied above, the thrombin receptor is in essence a peptide receptor; the agonist peptide bypasses thrombin-mediated receptor proteolysis and activates the thrombin receptor independent of thrombin and its protease activity. Agonist peptide caused both phosphoinositide hydrolysis and mitogenesis in CCL-39 cells. We next transfected CV-1 cells, which are normally poorly responsive to thrombin, with the platelet thrombin receptor cDNA. Expression of the cloned platelet thrombin receptor in CV-1 cells reproduced the responses normally seen in CCL-39 cells, both thrombin-induced phosphoinositide hydrolysis and mitogenesis. In both cell types, thrombin- and agonist peptide-induced phosphoinositide hydrolysis was largely pertussis toxin insensitive. Thus activation of two important thrombin-induced

events in CCL-39 fibroblasts occurs via the same unusual activation mechanism used for platelet activation, and very likely via the same or a very similar receptor.

Materials and Methods

Peptides

Agonist peptide SFLLRNPNDDKYEPF and control peptides LLRNPNDDKYEPF and FSLLRNPNDDKYEPF were synthesized by UCSF's Biomolecular Resource Facility (San Francisco, CA) or were a generous gift from Robert Scarborough, COR Therapeutics, Inc. (South San Francisco, CA). Peptides were purified by reverse phase HPLC before use.

Receptor-expressing Cell Lines

A cDNA encoding a functional human platelet thrombin receptor was subcloned into pBJ1, an expression vector derived from pcDL-SR α 296 (19) (pBJ1 was a generous gift from Dr. Mark Davis). The expression construct was cotransfected with a neomycin selection marker (18) into CV-1 cells by lipofection (6). Cells were grown in DME with 10% FCS, 100 U/ml penicillin, and 100 μ g/ml streptomycin. Stable transfectants were isolated using neomycin analogue G418 at concentrations of 0.8 mg/ml. Individual clones were selected and screened by dot blot for expression of thrombin receptor mRNA. The highest expressing clones were expanded and used in subsequent phosphoinositide turnover assays.

Phosphoinositide Hydrolysis Assays

Cells were seeded into 12-well plates at a density of 10^5 cells per well and grown to confluence. Wells were then rinsed once with 2 ml DME containing 25 mM Hepes, 100 U/ml penicillin, and 100 μ g/ml streptomycin. Cells were then incubated with 0.5 ml of DME containing 25 mM Hepes buffer, pH 7.4, 100 U/ml penicillin, and 100 μ g/ml streptomycin, and 2 μ Ci/ml [3 H]myo-inositol for 20–24 h at 37°C. Cells were treated with LiCl (20 mM final concentration) 1 min before addition of agonist. After 15 min of agonist treatment, all wells were rinsed once with 1 ml cold PBS, and cells were then extracted with 750 μ l 20 mM formic acid for 30 min at 4°C. In all experiments involving the use of Bordetella pertussis toxin islet-activating protein (List Biological Laboratories, Campbell, CA), cells were incubated at 37°C for 5 h before agonist addition in the presence of pertussis toxin at a final concentration of 100 ng/ml.

Cell extracts were loaded onto 1 ml packed volume columns of anion-exchange gel resin AGIX8, formate form, 100–200 mesh size (Bio-Rad Laboratories, Cambridge, MA) after columns were prepared with sequential washes of 2 ml 2 M ammonium formate/0.1 M formic acid, 2 ml water, and 4 ml 20 mM NH_4OH , pH 9.0. Immediately after loading, columns were washed with 3 ml 40 mM NH_4OH , pH 9.0, followed by 4 ml 40 mM ammonium formate. Columns were then eluted with 4 ml 2 M ammonium formate/0.1 M formic acid into scintillation vials containing 10 ml of scintillation cocktail and vials were counted in a scintillation counter (Beckman Instruments, Palo Alto, CA) for 10 min each. This procedure collects inositol mono, bis, and tris phosphates (11).

Mitogenesis Assays

CCL-39 cells and CV-1 cells were grown to confluence in 96 well plates (Costar, Cambridge, MA) in 100 μ l DME (Gibco Laboratories, Grand Island, NY) containing 10% FCS, 25 mM Hepes buffer, pH 7.4, 100 U/ml penicillin, and 100 μ g/ml streptomycin. Cells were then rinsed in DME and incubated in 100 μ l DME containing 25 mM Hepes buffer, pH 7.4, 100 U/ml penicillin, and 100 μ g/ml streptomycin for 48 h. Agonist was then added with or without 1 μ g/ml insulin, 5 μ g/ml transferrin, and 0.5 mg/ml BSA (see Fig. 2) and the incubation continued for 12 h. At this time [3 H]thymidine was added at 1.0 μ Ci/well (see Fig. 2) or 5.0 μ Ci/well. TCA-insoluble radioactivity was determined after an additional 24-h incubation.

Transfected and untransfected CV-1 cells (see Fig. 5) were treated with [3 H]thymidine (0.5 μ Ci/well) and agonists simultaneously and the plates incubated for an additional 48 h at 37°C. [3 H]thymidine incorporation into TCA-insoluble material was determined as previously described (5). [3 H]thymidine uptake results were confirmed by determining percent positive nuclei by [3 H]thymidine autoradiography as previously described (5).

ADP Ribosylation

Membrane preparation and ADP ribosylation was performed essentially as previously described (2). Thrombin receptor-transfected CV-1 cells were grown to confluence in eight 100-mm plates. Four of these plates were incubated in the presence of pertussis toxin (100 ng/ml final concentration) for 5 h at 37°C. Cells were then harvested using calcium/magnesium-free PBS with 0.4% EDTA, pelleted by centrifugation at 1,000 rpm for 10 min, resuspended in 800 μ l of lysis buffer (50 mM Tris-HCl, pH 7.5, 2.5 mM MgCl_2 , 1 mM EGTA, 1 mM PMSF, 1 mM DTT, and 1 mM benzamide-HCl) and lysed by freeze thawing and shearing through a 27-gauge needle. Cell nuclei were pelleted by centrifugation at 1,000 rpm for 5 min at 4°C and the membrane-containing supernatant was removed and centrifuged at 14,000 rpm for 20 min at 4°C. The membrane-containing pellet was resuspended in 200 μ l lysis buffer supplemented with 10% glycerol and protein concentrations were determined using Bradford reagent. 50 μ g of pertussis toxin-treated or -untreated membranes were suspended in 30 μ l of 50 mM Tris-HCl, pH 7.5, 1 mM EDTA, and 1 mM MgCl_2 and to each sample was added 50 μ l 2 \times reaction buffer (2 mM ATP, 100 mM Tris-HCl, pH 7.4, 20 mM thymidine, 40 mM arginine, 0.4 mg/ml BSA, 200 mM KPO_4 , pH 7.5, 10 mM ADP-ribose, 20 mM MgCl_2 , 2 mM EDTA, and 200 μ M GTP) and 4 μ l activated pertussis toxin (pertussis toxin was activated by incubating 20 μ l pertussis toxin [1 mg/ml] with 20 μ l 42 mM DTT, 20 mM Tris-HCl, pH 7.5 at 30°C for 20 min). Finally, 10 μ l of 5 μ M NAD containing 20 μ Ci [32 P]NAD was added to each sample and incubated at 30°C for 30 min. The reaction was stopped by adding 100 μ l of NAD wash solution (5 mM NAD, 50 mM Tris-HCl, pH 7.5, 25 mM EDTA) to each sample. Membranes were pelleted by centrifugation at 14,000 rpm for 5 min, washed twice with 200 μ l NAD wash solution, resuspended in 100 μ l Laemmli sample buffer, boiled for 5 min, and analyzed by SDS-PAGE (12% acrylamide gel) and autoradiograph. Exposure time was 5 h.

Results and Discussion

Agonist peptide caused phosphoinositide hydrolysis in CCL-39 cells with an EC_{50} of ~ 10 μ M (Fig. 1 A). The EC_{50} for thrombin-induced phosphoinositide hydrolysis in CCL-39 cells was ~ 30 pM (Fig. 1 B). The relative potencies of agonist peptide and thrombin in this system mimicked that seen for agonist-induced responses mediated by the cloned receptor in the oocyte system (22). Moreover, maximum responses elicited by saturating concentrations of agonist peptide (100 μ M) or thrombin (1 nM) were similar (Fig. 1, A and B). Thrombin receptor agonist peptide also stimulated [3 H]thymidine uptake in CCL-39 fibroblasts by approximately 20-fold (Fig. 2), and was once again several logs less potent than thrombin. The lower potency of agonist peptide compared to thrombin is predictable given the current model of receptor activation. Thrombin acts enzymatically; the peptide acts by binding. Thrombin generates a tethered ligand; the peptide is free. The kinetic advantages for thrombin are apparent.

The specificity of the agonist peptide was confirmed in several ways. First, a control peptide missing the agonist peptide's first two amino acids (see Materials and Methods) did not stimulate mitogenesis or phosphoinositide hydrolysis in CCL-39 cells (data not shown), even at concentrations as high as 100 μ M. Second, another control peptide in which the first two amino acids were switched in positions (SFLLRNPNDDKYEPF to FSLLRNPNDDKYEPF) had only minimal agonist activity (Fig. 2). Lastly, the actions of the agonist peptide were receptor dependent in the transfection studies discussed below.

The actions of the thrombin receptor agonist peptide described above strongly suggested that thrombin-induced phosphoinositide hydrolysis and mitogenesis in CCL-39

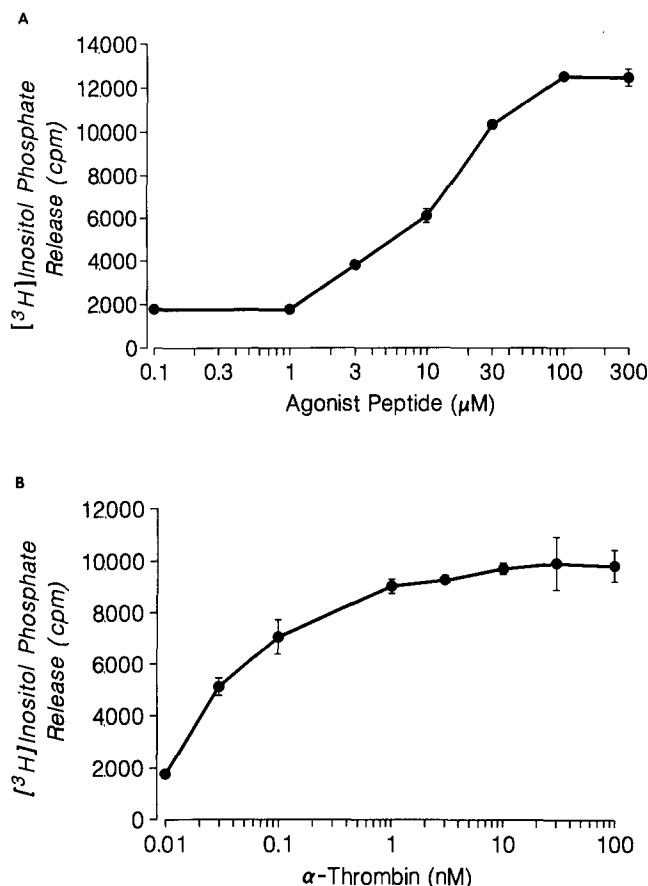


Figure 1. Thrombin-induced and thrombin receptor agonist peptide-induced phosphoinositide hydrolysis in CCL-39 fibroblasts. Quiesced CCL-39 fibroblasts loaded with [3 H]myo-inositol (see Materials and Methods) were treated in the presence of 20 mM LiCl for 15 min at 37°C with thrombin receptor agonist peptide (A) or α -thrombin (B) at the concentrations indicated. Total [3 H]inositol phosphates were collected and quantitated as described in Materials and Methods. Each point represents the mean of three replicate determinations with standard deviations as shown. These results are representative of three replicate experiments.

cells are mediated, at least in part, by the same novel proteolytic mechanism of receptor activation seen in platelets, probably by the same or closely related receptors. Northern analysis of mRNA from CCL-39 fibroblasts suggested that the hamster homologue of the cloned human platelet thrombin receptor was expressed in this hamster fibroblast cell line (Fig. 3) as recently reported (12). Interestingly, Pouyssegur and colleagues have reported that thrombin receptor agonist peptide elicited robust mitogenic effects on their subclone of CCL-39 cells only when FGF was included in the medium, while thrombin itself was mitogenic in the absence of FGF (21). We have not observed a requirement for additional mitogens for the action of agonist peptide on the CCL-39 cells used in our laboratory (Fig. 2). We do wish to state, however, that our data in no way preclude the existence of other thrombin receptors.

To determine whether the cloned human platelet thrombin receptor could itself account for thrombin-induced cellular events in non-platelet cells, thrombin receptor cDNA was stably transfected into CV-1 cells and the ability of receptor-transfected cells to respond to both α -thrombin and throm-

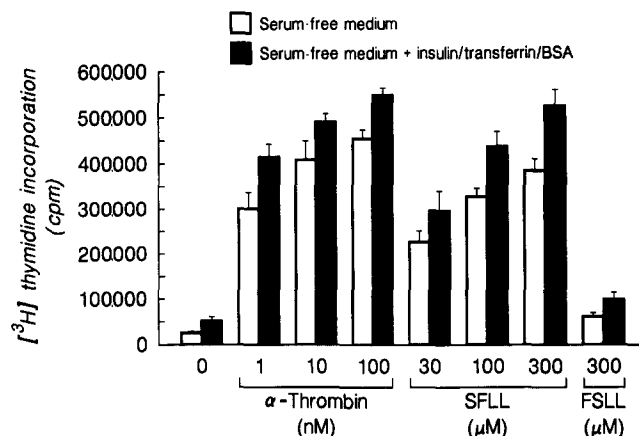


Figure 2. Thrombin and thrombin receptor agonist peptide cause mitogenesis in CCL-39 fibroblasts. CCL-39 fibroblasts were incubated in either serum-free DME (open columns) or serum-free DME with 1 μ g/ml insulin, 5 μ g/ml transferrin, and 0.5 mg/ml BSA (closed columns) for 48 h at 37°C. Cells were then stimulated with either α -thrombin, thrombin receptor agonist peptide (SFLLRNPNPKYEPEF; SFL), or control peptide (FSLLRNPNPKYEPEF; FSLL) at the concentrations indicated for 12 h at 37°C, followed by addition of [3 H]thymidine in the continued presence of agonists for an additional 24 h at 37°C. [3 H]thymidine incorporation into TCA-insoluble material was determined as previously described (5). Each point represents the mean of three replicate determinations with standard deviations as shown. These results were replicated in three separate experiments. Similar results were obtained when the [3 H]thymidine concentration was increased five-fold, such that [3 H]thymidine could not be limiting.

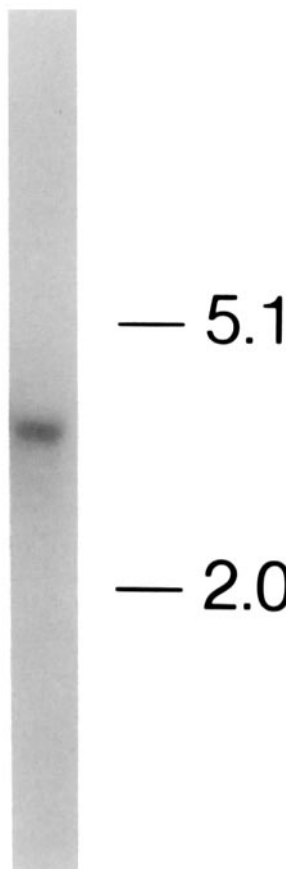


Figure 3. Detection of thrombin receptor mRNA in CCL-39 fibroblasts. Five micrograms of poly(A)⁺ RNA from CCL-39 fibroblasts was denatured in glyoxal-DMSO and subjected to Northern analysis using standard techniques (13). The blot was hybridized at high stringency with random primer-generated 32 P-labeled probe representing the entire coding region of the cloned human platelet thrombin receptor; hybridized probe was detected by autoradiography. The migration of 28S and 18S RNA is shown at right.

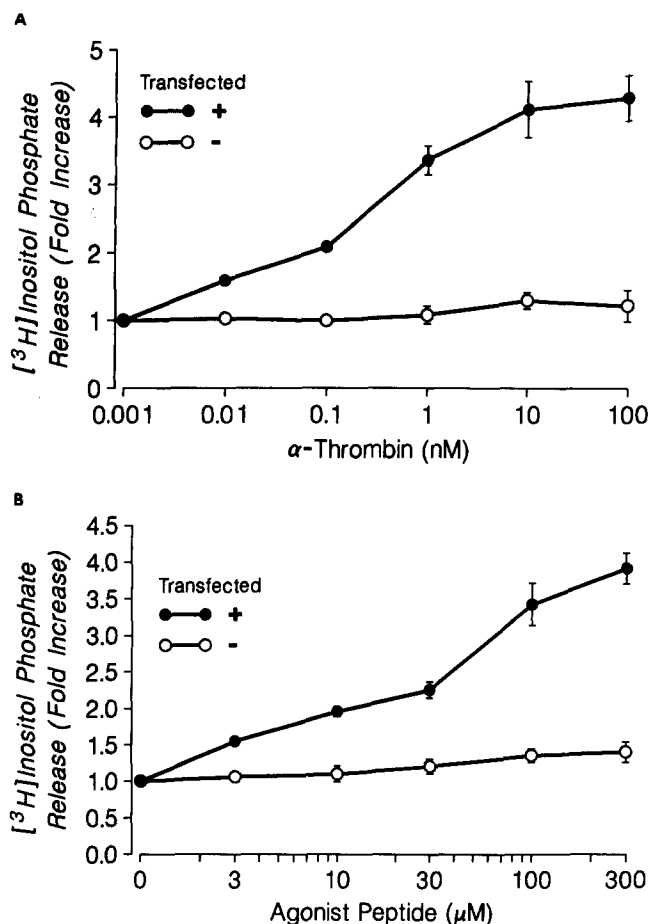


Figure 4. Transfection with platelet thrombin receptor cDNA confers thrombin-induced and thrombin receptor agonist peptide-induced phosphoinositide hydrolysis on CV-1 cells. Thrombin receptor-transfected (●) and untransfected (○) CV-1 cells labeled with [3 H]myo-inositol (see Materials and Methods) were treated in the presence of 20 mM LiCl for 15 min at 37°C with α -thrombin (A) or thrombin receptor agonist peptide (B) at the concentrations indicated. Total [3 H]inositol phosphates were collected and quantitated as described in Materials and Methods. [3 H]inositol phosphate release induced by 0.001 nM α -thrombin was equivalent to baseline (i.e., did not stimulate inositol phosphate release). Each point represents the mean of three replicate determinations with standard deviations as shown. These results are representative of three replicate experiments.

bin receptor agonist peptide was examined. CV-1 cells transfected with the human platelet thrombin receptor responded with a 400% increase in inositol phosphate release when stimulated with saturating concentrations of either thrombin or agonist peptide (Fig. 4, A and B). The EC_{50} 's for thrombin and thrombin receptor agonist peptide were 100 pM and 10 μ M, respectively, similar to the EC_{50} 's found in naturally thrombin-responsive CCL-39 cells and in oocytes expressing the platelet thrombin receptor cDNA (22). By contrast, untransfected CV-1 cells exhibited no significant increase in phosphoinositide hydrolysis when stimulated with either thrombin or thrombin receptor agonist peptide at concentrations as high as 100 nM and 300 μ M, respectively (Fig. 4, A and B). Thrombin failed to activate phosphoinositide turnover in other clones expressing thrombin receptor mutants rendered unactivatable by cleavage site mutations (Hung,

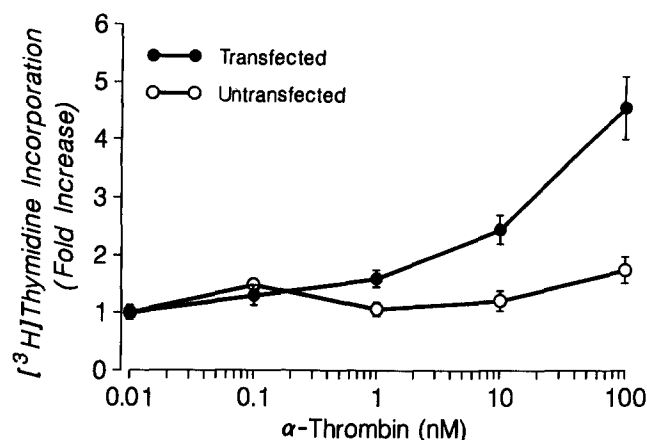


Figure 5. Transfection with platelet thrombin receptor cDNA confers thrombin-induced and thrombin receptor agonist peptide-induced mitogenesis on CV-1 cells. Quiesced thrombin receptor-transfected (●) and untransfected (○) CV-1 cells were stimulated with the indicated concentrations of α -thrombin for 48 h at 37°C and [3 H]thymidine incorporation into TCA-insoluble material was determined as previously described (5). Mean cpm per well of unstimulated receptor-transfected and -untransfected cells were similar (5,940 \pm 738 [S.E.] and 6,566 \pm 1,447 [S.E.], respectively).

D. T., T.-K. H. Vu, and S. R. Coughlin, manuscript in preparation). While anticipated from our previous studies (22), this is the first demonstration that the cloned thrombin receptor couples to phosphoinositide hydrolysis.

Pertussis toxin pretreatment

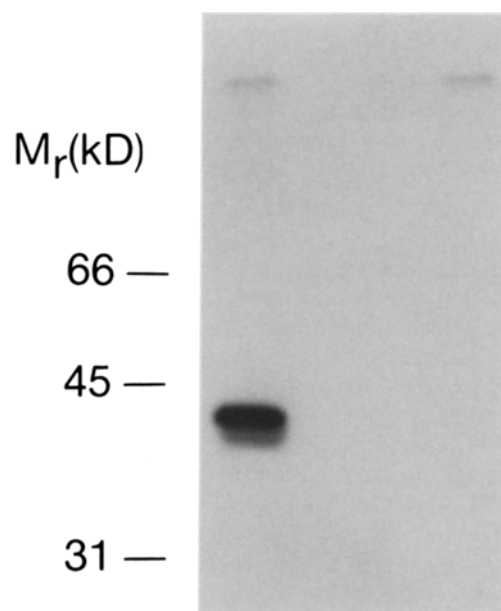


Figure 6. Demonstration of a lack of pertussis substrate in membranes from pertussis toxin-treated cells. Membranes from pertussis toxin-treated (+) or untreated (-) thrombin receptor-transfected CV-1 cells were analyzed for available pertussis substrates as described in Materials and Methods. Quantitation of radioactivity in the + lane band and in the corresponding region of the - lane revealed that at least 97% of pertussis substrate had been "removed" by pertussis toxin pretreatment.

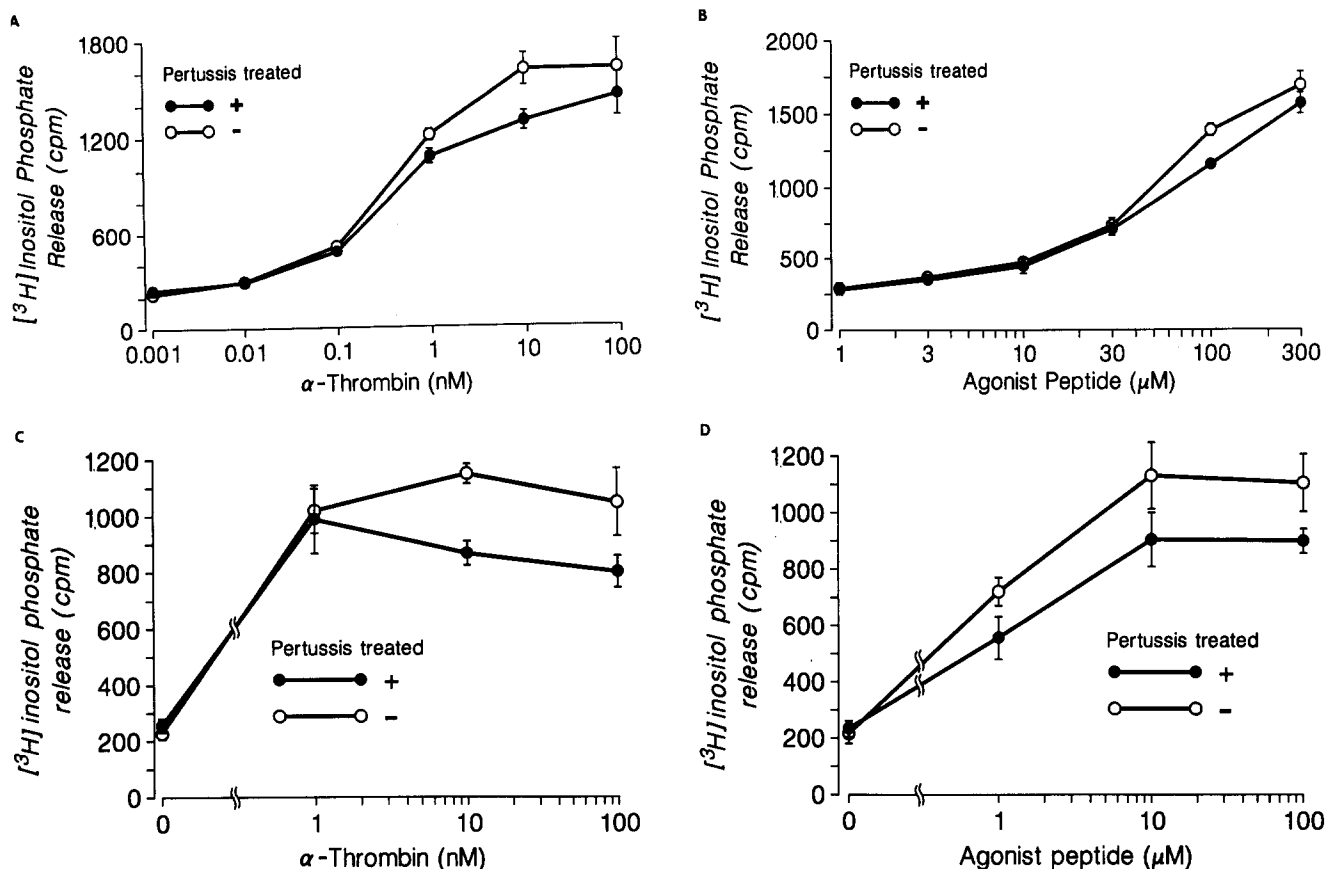


Figure 7. Thrombin-induced and thrombin receptor agonist peptide-induced phosphoinositide hydrolysis in thrombin receptor-transfected CV-1 cells and CCL-39 fibroblasts is largely pertussis toxin-insensitive. Quiescent thrombin receptor-transfected CV-1 cells (A and B) or CCL-39 fibroblasts (C and D) labeled with [3 H]myo-inositol (see Materials and Methods) were incubated at 37°C for 5 h with either 10 μ l of PBS (○) or an equal volume of PBS containing pertussis toxin to yield a final concentration 100 ng/ml (●). Cells were then treated for 15 minutes at 37°C with α -thrombin (A and C) and thrombin receptor agonist peptide (B and D) at the concentrations indicated in the presence of 20 mM LiCl. Total [3 H]inositol phosphates were collected and quantitated as described in Materials and Methods. Each point represents the mean of three replicate determinations with standard deviations as shown. These results are representative of three replicate experiments.

The cloned "platelet thrombin receptor" also conferred thrombin-induced mitogenesis to CV-1 cells. [3 H]thymidine uptake in CV-1 cells stably transfected with the cloned thrombin receptor cDNA increased ~500% in response to thrombin, while untransfected CV-1 cells showed minimal response (Fig. 5).

These studies show that transfection of CV-1 cells with the cloned platelet thrombin receptor cDNA conferred the thrombin responses seen in naturally thrombin-responsive CCL-39 cells. The similarities in the thrombin and peptide responses in native CCL-39 cells and receptor-transfected CV-1 cells extended to their pertussis toxin sensitivity. Pertussis toxin treatment sufficient to fully ADP-ribosylate available G-protein α subunits (Fig. 6) had little inhibitory effect on thrombin- or agonist peptide-stimulated phosphoinositide hydrolysis in thrombin receptor-transfected CV-1 cells (Fig. 7, A and B). In CCL-39 cells, pertussis toxin treatment caused only partial inhibition of thrombin- or agonist peptide-stimulated phosphoinositide hydrolysis (Figure 7, C and D). Published studies report both pertussis-sensitive and -insensitive thrombin-induced phosphoinositide hydrolysis in various cell lines (7, 9, 11, 14, 15), suggesting that the thrombin receptor may couple to phosphoinositide hydrolysis via different G-proteins determined by a cell's G-protein repertoire,

or that different receptors may exist. Our studies demonstrate that the cloned thrombin receptor couples to phosphoinositide hydrolysis predominantly via a pertussis toxin-insensitive G-protein(s) in the cell lines we examined. The recently described G_q class of G-proteins, which lack a "pertussis site" (17) and couple to the β 1 isozyme of phospholipase C (20), are candidates for mediating thrombin receptor-induced phosphoinositide hydrolysis.

In summary, these studies suggest that the unique mechanism of thrombin receptor activation described for platelet activation also mediates important thrombin-induced events (phosphoinositide hydrolysis and mitogenesis) in a naturally thrombin-responsive fibroblast cell line. Moreover, our studies show that the platelet thrombin receptor can confer these same responses to a cell line that is normally poorly responsive to thrombin. Our results strongly suggest that the hamster homologue of the cloned human "platelet thrombin receptor" accounts, at least in part, for thrombin responses in CCL-39 fibroblasts and that the cloned platelet thrombin receptor may mediate not only platelet activation but also mitogenesis and other thrombin-induced events in a variety of cell types. In this context, we have recently found thrombin receptor mRNA expressed in vascular endothelium and smooth muscle cells by PCR and Northern analysis (22, and

data not shown) and by in situ hybridization of human arteries (data not shown). Thus antagonists of the cloned thrombin receptor may be useful not only in inhibiting platelets and thrombosis, but also in blocking undesirable thrombin-induced inflammatory or proliferative responses in the blood vessel wall.

We thank Dr. John Fenton II (Wadsworth Center/Albany Medical College, Albany, NY) for his generous gift of α -thrombin and Dr. Mark Davis (Stanford Medical Center, Stanford, CA) for his generous gift of the expression plasmid pBJ1 used in these studies. We also thank Drs. Henry R. Bourne and Yung H. Wong for valuable discussions.

This work was supported in part by the Physician Scientist Award National Institutes of Health HL02487 (D. T. Hung), National Institutes of Health HL44907 and HL43801 (S. R. Coughlin), Pacific Vascular Foundation (N. A. Nelken) and by the University of California Tobacco-Related Disease Research Program 2RT19. This work was performed during the tenure of an award from the American Heart Association and SmithKline Beecham (S. R. Coughlin).

Received for publication 7 August 1991 and in revised form 17 October 1991.

References

1. Bar-Shavit, R., A. Kahn, G. D. Wilner, and J. W. Fenton II. 1983. Monocyte chemotaxis: stimulation by specific exosite region in thrombin. *Science (Wash. DC)*. 220:728-731.
2. Chang, F.-H., and H. R. Bourne. 1987. Dexamethasone increases adenyl cyclase activity and expression of the α -subunit of G_s in GH₃ cells. *Endocrinology*. 121:1711-1715.
3. Chen, L. B., and J. M. Buchanan. 1975. Mitogenic activity of blood components. I. Thrombin and prothrombin. *Proc. Natl. Acad. Sci. USA*. 72:131-135.
4. Chen, L. B., N. N. H. Teng, and J. M. Buchanan. 1976. Mitogenicity of thrombin and surface alterations on mouse splenocytes. *Exp. Cell Res.* 101:41-46.
5. Coughlin, S. R., W. M. F. Lee, P. W. Williams, T. M. Giels, and L. T. Williams. 1985. C-myc gene expression is stimulated by agents that activate protein kinase C and does not account for the mitogenic effect of PDGF. *Cell*. 43:243-251.
6. Felgner, P. L., T. R. Gadek, M. Holm, R. Roman, H. W. Chan, M. Wenz, J. P. Northrop, G. M. Ringold, and M. Danielsen. 1987. Lipofection: a highly efficient, lipid-mediated DNA-transfection procedure. *Proc. Natl. Acad. Sci. USA*. 84:7413-7417.
7. Garcia, J. G. N., R. G. Painter, J. W. Fenton II, D. English, and K. S. Callahan. 1990. Thrombin-induced human endothelial cell PGI₂ biosynthesis in human endothelium: role of guanine nucleotide regulatory proteins in stimulus/coupling responses. *J. Cell. Physiol.* 142:186-193.
8. Hanson, S. R., and L. A. Harker. 1988. Interruption of acute platelet-dependent thrombosis by the synthetic antithrombin PPACK. *Proc. Natl. Acad. Sci. USA*. 85:3184-3188.
9. Jones, L. G., P. M. McDonough, and J. H. Brown. 1989. Thrombin and trypsin act at the same site to stimulate phosphoinositide hydrolysis and calcium mobilization. *Molecular Pharmacology*. 36:142-149.
10. Liu, L.-W., T.-K. H. Vu, C. T. Esmon, and S. R. Coughlin. 1991. The region of the thrombin receptor resembling hirudin binds to thrombin and alters enzymes specificity. *J. Biol. Chem.* 266:16977-16980.
11. Paris, S., and J. Pouyssegur. 1986. Pertussis toxin inhibits thrombin-induced activation of phosphoinositide hydrolysis and Na⁺/H⁺ exchange in hamster fibroblasts. *EMBO (Eur. Mol. Biol. Organ.) J.* 5(1):55-60.
12. Rasmussen, U. B., V. Vouret-Craviari, S. Jallat, Y. Schlesinger, G. Pages, A. Pavirani, J.-P. Lecocq, J. Pouyssegur, and E. Van Obberghen-Schilling. 1991. cDNA cloning and expression of a hamster α -thrombin receptor coupled to Ca²⁺ mobilization. *FEBS (Fed. Eur. Biol. Soc.) Lett.* 288(1,2):123-128.
13. Sambrook, J., E. F. Fritsch, and T. Maniatis. 1989. In C. Nolan, editor. Molecular cloning. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York.
14. Seuwen, K., A. Lagarde, and J. Pouyssegur. 1990. Deregulation of hamster fibroblast proliferation of mutated ras oncogenes is not mediated by constitutive activation of phosphoinositide-specific phospholipase C. *EMBO (Eur. Mol. Biol. Organ.) J.* 7(1):161-168.
15. Seuwen, K., C. Kahan, T. Hartmann, and J. Pouyssegur. 1990. Strong and persistent activation of inositol lipid breakdown induces early mitogenic events but not G₀ to S phase progression in hamster fibroblasts. *J. Biol. Chem.* 265(36):22292-22299.
16. Shuman, M. A. 1986. Thrombin-cellular interactions, p. 228-239. In D. A. Walz, J. W. Fenton II, and M. A. Shuman, editors. Bioregulatory functions of thrombin. Vol. 485. New York Academy of Sciences, New York.
17. Strathmann, M., and M. I. Simon. 1990. G protein diversity: a distinct class of α subunits is present in vertebrates and invertebrates. *Proc. Natl. Acad. Sci. USA*. 87:9113-9117.
18. Southern, P. J., and P. Berg. 1982. Transformation of mammalian cells to antibiotic resistance with a bacterial gene under control of the SV40 early region promoter. *J. Mol. Appl. Genet.* 1:327-336.
19. Takebe, Y., M. Seiki, J.-I. Fujisawa, P. Hoy, K. Yokota, K.-I. Arai, M. Yoshida, and N. Arai. 1988. SR α promoter: an efficient and versatile mammalian cDNA expression system composed of the simian virus 40 early promoter and the R-U5 segment of human T-cell leukemia virus type I long terminal repeat. *Mol. Cell. Biol.* 8(1):466-472.
20. Taylor, S. J., H. Z. Chae, S. G. Rhee, and J. H. Exton. 1991. Activation of the β 1 Isozyme of phospholipase C by the α subunits of the G_q class of G proteins. *Nature (Lond.)*. 350(11):516-518.
21. Vouret-Craviari, V., E. Van Obberghen-Schilling, U. B. Rasmussen, A. Pavirani, J.-P. Lecocq, and J. Pouyssegur. 1992. Synthetic α -thrombin receptor peptides activate G protein-coupled signalling pathways but are unable to induce mitogenesis. *Mol. Biol. Cell*. In press.
22. Vu, T.-K. H., D. T. Hung, V. I. Wheaton, and S. R. Coughlin. 1991. Molecular cloning of a functional thrombin receptor reveals a novel proteolytic mechanism of receptor activation. *Cell*. 64:1057-1068.
23. Vu, T.-K. H., V. I. Wheaton, D. T. Hung, I. Charo, and S. R. Coughlin. 1991. Domains specifying thrombin-receptor interaction. *Nature (Lond.)*. 353:674-677.